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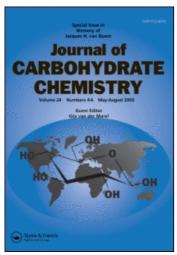
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A SIMPLE ROUTE TO β.β-TREHALOSE VIA TRICHLOROACETIMIDATES

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ABSTRACT

The reaction of 2,3,4,6-tetra-0-acetyl- β -D-glucopyranose with 2,3,4,6-tetra-0-acetyl- α -D-glucopyranosyl trichloroacetimidate in CH₂Cl₂ promoted by BF₃;0Et₂ gives β , β -octaacetyltrehalose in up to 58% isolated yield, which is readily deacetylated to β , β -trehalose. The corresponding 2,3,4,6-tetra-0-acetyl- β -D-glucopyranosyl pentafluorophenylimidate is configurationally stable and inert to coupling under mild conditions.

INTRODUCTION

The ready availability of α, α -trehalose from natural sources, and the interest in some of its biologically active derivatives has led to extensive development of its chemistry and the preparation of a range of derivatives. By contrast, the anomer β, β -trehalose, which does not occur naturally, has not been studied to the same extent, and has not been readily obtainable in pure form. Conventional variants on the Koenigs-Knorr reaction give low yields of β, β -trehalose derivatives and require tedious separations of the product from its stereoisomers. This makes the imidate coupling route, whose most

$$AcOH_2C$$
 OAC OAC

<u>Scheme</u>

effective practical variant is the $\beta\text{-specific}$ acid-catalysed reaction of alcohols with $\alpha\text{-anomeric}$ trichloroacetimidates, 4 particularly attractive. We show that this can be applied to the synthesis of octaacetyl β , β -trehalose in up to 58% isolated yield, according to the Scheme.

RESULTS AND DISCUSSION

There have been few reports in the literature on the chemistry of the interesting disaccharide β because the conventional syntheses use unpleasant reagents such as $\mathrm{Hg}(\mathsf{CN})_2$ or require tedious chromatographic separations. Formation of glycosidic linkages by the trichloroacetimidate variant of the imidate coupling reaction has now been applied to β be trehalose synthesis. Reaction occurs under mild acid catalysis and the co-product is trichloroacetamide. This makes the method very suitable for large-scale preparative work, and base-catalysed methanolysis of the intermediate β be trehalose octaacetate leads directly to the crystalline disaccharide. The $^1\mathrm{H}$ NMR spectrum of product in $\mathrm{D}_2\mathrm{O}$ shows a number of differences from that of the α a-anomer which are tabulated below. Axial ring-protons at C2, C3 and C5 are more shielded by 0.23 to 0.36 ppm in the β because

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TABLE Chemical shifts of trehalose protons in D₂O, relative to internal DSS. The spectra were recorded using a Bruker WH 300 machine operating at 24°C with a sweep width of 4KHz, pulse width 60° and 32K data points. Coupling constants, β , β anomer: J_{1,2} = 7.5; J_{2,3} = 9.2; J_{3,4} = 8.9; J_{4,5} = 9.5; J_{5,6a} = 2.2; J_{5,6b} = 5.6; J_{6a,6b} = 12.4 Hz. We thank Dr. S.J. Kimber for these measurements.

Proton	H ₁	H ₂	H 3	H4	H ₅	H ₆ a	Heb
ββ	4-74	3-35	3-45	3-29	3-40	3-85	3-67
αα	5-20	3-64	3-84	3-44	3-81	3-80	3-77

It was felt that the corresponding pentafluorophenylimidate would provide a useful alternative starting material since 2,3,4,6-tetra--0-acetyl- α -0-glucopyranosyl trichloroacetimidate is a viscous oil and trace impurities were evident in the ^1H NMR spectrum. Replacement of CCl₃CN with C₆F₅CN in the imidate synthesis did indeed lead to a crystalline product but we were surprised to discover that this was the pure β -anomer. All attempts to equilibrate it with the (presumed more stable) α -anomer using KH in CH₂Cl₂, a silica gel, led to its quantitative recovery. An example of the "reverse anomeric effect" cannot be ruled out. 6

EXPERIMENTAL

General Methods

¹H-Nuclear magnetic resonance spectra were obtained on a Bruker WH 300 (300.13 MHz) instrument. Chemical shifts (δ) are expressed in parts per million from tetramethylsilane. The mass spectrum was recorded on a V.G. Micromass spectrometer operating in electron impact mode. The infra-red spectrum was recorded on a Unicam SP 1000 spectrometer as a nujol mull. Optical rotations were measured on a Perkin-Elmer 141 polarimeter.

Commercial solvents were distilled prior to use from an appropriate drying agent according to standard procedures. Dichloromethane was distilled from phosphorus pentoxide; diethyl ether was distilled from sodium wire employing sodium benzophenone ketyl as an indicator; methanol was distilled from magnesium turnings.

2.3.4.6-Tetra-O-acetyl- α -D-glucopyranosyl trichloro-acetimidate. To a stirred, argon-blanketed solution of 2.3.4.6-tetra-O-acetyl- β -D-glucose (1.50 g, 4.12 mmol) and distilled CCl₃CN 1.50 ml, 2.16 g., 15 mmol) in dry CH₂Cl₂(10 ml) at -78°, there was added a suspension of potassium hydride 0.20 g, 4.99 mmol) in dry CH₂Cl₂ (10 ml). The suspension was allowed to warm to ambient temperature over 0.5 h. The potassium salts then filtered off and the solution was passed through a short column of silica gel and concentrated in vacuo. 2.3.4.6-Tetra-O-acetyl- α -D-glucopyranosyl trichloroacetimidate was obtained as an oil (1.6 g, 76%). ¹H NMR [CDCl₃,300 MHz]: δ 8.7(brs,NH) δ .55(d,H₁,J_{1,2} = 4 Hz) 5.55 t.H₃,J_{2,3} = J_{3,4} = 9.5 Hz) 5.2(t,H₄,J_{4,5} = 9.5 Hz) 5.15(dd,H₂ J_{1,2} = 4 Hz) 4.25(m,H₅,H₆b) 4.15(dd,H_{6a} J_{6,6} = 12.5 Hz,J_{5,6a} = 2.5 Hz) 2.0 - 2.15 (4 x S,COCH₃) p.p.m.

2.3.4.6-Tetra-O-acetyl β-D-glucopyranosyl pentafluorophenylimidate. An argon-blanketed solution of 2,3,4,6-tetra-0--acetyl-β-D-glucose (0.90 g. 2.47 mmol) and pentafluorobenzonitrile 0.80 ml, 1.22 g, 6.6 mmol) in dry CH_2Cl_2 (20 ml) at -78° , was added to dry potassium hydride (0.13 g, 3.24 mmol) also held at -78° . The mixture was stirred at -78° for 0.25 h and then warmed to ambient temperature over 0.5 h. The reaction mixture was passed through a short column of silica gel and concentrated in vacuo. Recrystallization of the resulting solid from CH₂Cl₂/Et₂O gave white crystals of 2.3.4.6-tetra-0-acetyl-£-D-glucopyranosyl pentafluorophenylimidate (1.24 g, 93%) m.p. $139.5-42^{\circ}$. $[\alpha]_{D}^{20} = -13.7^{\circ}$ (c,4.8,CH₂Cl₂). Found: C.47.13; H.3.76; C21H20F5NO10 requires: C.46.59; H.3.72%. M.S. m/z 539 $(M^{+}-2, 67\%)$ 331 $(M^{+}-C_{6}F_{5}CONH, 100\%)$ 213 $(C_{6}F_{5}C^{+}(OH)NH_{2}, 26\%)$ ¹H NMR [(CD₃)₂SO.300 MHz] δ 5.7 (d.H₁,J_{1,2} = 8 Hz) 5.45(t.H₃,J_{2,3} = J_{3,4} = 9.5 Hz) 5.13(dd, H_2) 5.03(t, H_4 , $J_{4,5}$ = 10 Hz) 4.2(dd, H_{6a} , $J_{6,6}$ = 12.5 Hz, $J_{6a,5}$ = 5 Hz) 4.1(brm, H₅) 4.02(dd, H₆b, J₆b, $_5$ = 2 Hz) 2.1 = 2.0 (4 x s, COCH₃)ppm. In CDCl $_3$ the $^1\mathrm{H}$ NMR spectrum was highly second-order in the 5.2 ppm region IR (nujol) 1745(s) 1230(s) cm⁻¹.

 $\frac{2.3.4.6-\text{Tetra-0-acetyl-}\beta-\underline{D-glucopyranosyl\ 2'.3'.4'.6'-}{-\text{tetra-0-acetyl-}\beta-\underline{D-glucopyranoside;}\ (\beta.\beta-\text{trehalose\ octaacetate}).}$ To a mixture of 1.14 g (2.24 mmol) 2.3.4.6-tetra-0-acetyl- α -D-glucopyranosyl trichloroacetimidate and 0.816 g (2.24 mmol)

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2,3,4,6-tetra-0-acetyl-β-D-glucose (freshly prepared) in CH₂Cl₂ (25 ml) at 0° , was added 0.6 ml (4.88 mmol) BF₃: Et₂0 in CH2Cl2 (5 ml) with stirring. The solution stood for 16 h at room temperature, then saturated NaHCO3 aq. (15 ml) was added and the separated organic phase washed with water $(3 \times 15 \text{ ml})$ and saturated NaCl solution, dried over Na₂SO₄, and then solvent removed in vacuo. Recrystallization from Et₂0/40-60 petrol gave 2,3,4,6-tetra-0-acetyl-β-D-glucopyranosyl 2',3',4',6'--tetra-0-acetyl- β - \underline{D} -glucopyranoside (0.666 g). Further product was obtained by flash chromatography (Et₂0/40-60 petrol, 9:1) and combined to give 0.869 g (55%) of white needles, m.p. 181.5 - $183^{\circ}(\text{lit.}^{2} 181.5 - 182.5^{\circ}); [\alpha]_{0}^{20} -15.5^{\circ}(c5.4, \text{CHCl}_{3}), \text{lit.}^{3} -17^{\circ}(\text{CHCl}_{3});$ ¹H NMR [CDCl₃, 300 MHz] 5.2 (t,H₃,J_{2,3} = J_{3,4} = 8.5 Hz) 5.15 $(t_1H_4,J_{4,5} = 9 \text{ Hz}) 5.0(t_1H_2,J_{1,2} = 8.5 \text{ Hz}) 4.9(d_1H_1,J_{1,2} = 8 \text{ Hz})$ $4.25(dd.H_{6a}, J_{5,6} = 4.5 Hz, J_{6,6} = 12.5 Hz) 4.15(dd.H_{6b}, J_{5,6b} =$ 2.5 Hz) 3.75(ddd, H_5) 2.0 = 2.25(4 x s, COCH₃) ppm.

A large scale preparation afforded 16.91 g (62%) of crude material which gave 15.82 g (58%) of pure material on recrystallization as before, without the requirement for chromatography.

<u>β-D-Glucopyranosyl</u> <u>β-D-glucopyranoside</u> (β,β-trehalose). Methanolic NaOMe (1.45 ml 0.1 M) was added to a solution of 2.3.4.6-tetra-0-acetyl-β-D-glucopyranosyl 2'.3'.4'.6'-tetra-0-acetyl-β-D-glucopyranoside (2.24 g, 3.16 mmol) in hot MeOH. The solution was then refluxed for 0.25 h, solvent removed in vacuo and the solid was precipitated from MeOH/EtOH and dried in vacuo at 64° for 3 h and then 110° for 14 h. There was thus obtained β-D-glucopyranosyl β-D-glucopyranose as an off-white solid (1.05g, 97%) m.p. 129-34° ([it³/₂ 135-40°), [α] $^{20}_{D}$ = -35.2°(c5.0,H₂0), lit -40°(H₂0). A single recrystallization from H₂0/Me₂CO gave colourless plates, m.p. 134-9°, [α] $^{20}_{D}$ = -39°(7.4,H₂0); ¹H NMR [D₂0,300 MHz] recorded in Table.

Attempted epimerisation of 2.3.4.6-tetra-0-acetyl- β -D-glucopyranosyl pentafluorophenylimidate

To a solution of 2,3,4,6-tetra-0-acetyl- β -D-glucopyranosyl pentafluorophenylimidate (0.63 g,1.17 mmol) in CH₂Cl₂(12 ml) was added distilled boron trifluoride etherate (0.15 ml,1.2 mmol).

The solution was stirred for 24 h at room temperature, then saturated aqueous $NaHCO_3$ (10 ml) was added and the separated organic phase washed with water (3 x 5 ml), dried (anhydrous magnesium sulphate) and the solvent was removed <u>in vacuo</u>. Examination of the residue (^1H-NMR) showed only the presence of unreacted starting material.

Attempted coupling of 2,3,4,6-tetra-0-acetyl- β -D-glucopyranosyl pentafluorophenylimidate and methanol

To a solution of 2,3,4,6-tetra-0-acetyl- β -D-glucopyranosyl pentafluorophenylimidate (0.15 g,0.28 mmol) and methanol (0.050 ml, 1.2 mmol) in CH₂Cl₂ (5 ml) was added distilled boron trifluoride etherate (0.040 ml, 0.32 mmol) at 0°. The solution was stirred for 16 h at room temperature and the solvent was then removed in vacuo. Examination of the residue (1 H-NMR) showed only the presence of unreacted starting material.

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